

Role of NAD(P) H:quinone oxidoreductase polymorphism at codon 187 in susceptibility to lung, laryngeal and oral/pharyngeal cancers

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NAD(P)H:quinone oxidoreductase (NQO1) has been proposed to play a protective role against the toxic effects of benzo[a]pyrene quinones. The C^{609} T base change in the NQOI gene, resulting in a Pro¹⁸⁷Ser amino acid change in the protein, has been associated with deficient enzyme activity. We examined whether this polymorphism modified the risks of smoking-related cancers in a case-control study involving patients with lung cancer (n = 150), laryngeal cancer (n = 129), oral/pharyngeal cancer (n = 121) and control individuals (n = 172), all Caucasian smokers. No statistically significant associations were observed between the NQO1 genotypes and smoking-related cancers, although the Ser/ Ser genotype was associated with a tendency towards increased risk for lung cancer (odds ratio [OR] = 2.2, 95% confidence interval [CI] 0.7-6.7) and for oral/pharyngeal cancer (OR = 2.3, 95% CI 0.6-8.2). No significant interaction between the NQOI genotype and either smoking exposure or GSTM1 genotype was found. Our results are consistent with the hypothesis that lack of NQO1 activity may be involved in some smoking-related cancers. However, they were based on small numbers of individuals with the putative atrisk genotype, and the associations did not reach statistical significance. Moreover, these results contrast with those observed in some other ethnic populations, where a protective effect of the NOO1 Ser allele was found. Further studies are therefore clearly needed for a better understanding of the potential role of NQO1 activity in tobacco-related cancers.

Keywords: lung cancer, laryngeal cancer, oral cavity/pharyngeal cancer, NQO1 genotype, epidemiology, tobacco smoking

Introduction

Several tobacco carcinogens are metabolized via complex enzymatic mechanisms. Benzo[a]pyrene, a major constituent of tobacco smoke, undergoes oxidative metabolism to produce highly reactive benzo[a]pyrene quinones (BPQs) (Workman 1994), which are further metabolized by competing enzymes. A one-electron reduction of BPQs by NADPH:cytochrome P450 reductase results in the formation of semiquinones and reactive oxygen species, leading to cellular damage (Joseph and Jaiswal 1994). On the other hand, the two-electron-reducing NAD(P)H:quinone oxidoreductase 1 (NQO1) catalyses the conversion of quinones

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to relatively stable hydroquinones, which are readily excreted from the cells after conjugation (Talalay et al. 1995). NQO1 has been shown to specifically prevent the formation of BPQ-DNA adducts generated by CYP1A1 and P450 reductase (Joseph and Jaiswal 1994). A C to T base substitution at position 609 of the NOO1 cDNA results in a proline (Pro) to serine (Ser) amino acid change at position 187 of the protein (Traver et al. 1992, 1997, Roswold et al. 1995). NQO1 is expressed in normal lung tissue but is undetectable in lungs from individuals homozygous for the NOO1 Ser allele (Siegel et al. 1999). NOO1 polymorphism may therefore be a particularly important modifier of susceptibility to smokingrelated cancers.

There are only a few previous studies on the possible association between NQO1 genotypes and lung cancer, and these have yielded inconclusive results (Roswold et al. 1995, Wiencke et al. 1997, Chen et al. 1999, Lin et al. 1999). Moreover, the relationship between NQO1 polymorphism and cancers of the upper aerodigestive tract has not been previously examined.

We have conducted a hospital-based case-control study in France to investigate the possible role of several polymorphic genes of xenobiotic-metabolizing enzymes in the aetiology of smoking-related cancers among Caucasian smokers. Here, we extended the study to examine whether the NQO1 genotypes modify the risks of lung, laryngeal and oral cavity/pharyngeal cancers, either alone or in combination with glutathione S-transferase M1 (GSTM1), which is known to be involved in detoxification of reactive metabolites of carcinogenic substances from tobacco smoke. Studies on lung cancer have suggested an association with the GSTM1 null genotype (Vineis et al. 1999). We also investigated the potential modifying role of smoking exposure in the relationships between cancer risks and NQO1 genotypes.

Materials and methods

Study population

The study subjects were recruited between 1988 and 1992 in 10 French hospitals, nine of which are located in Paris (Bouchardy et al. 1997, Jourenkova-Mironova et al. 1999). Cases were all Caucasian patients with histologically confirmed incident squamous cell carcinomas of the lung, larynx, oral cavity or pharynx. Patients with small cell carcinomas of the lung were also eligible. A control group, frequency matched on age, sex and hospital, consisted of Caucasian patients without previous or current malignant disease. All cases and controls had to be regular smokers, defined as individuals having smoked five cigarettes or more (or cigars or pipe) per day for at least 5 years. Subjects were recruited by seven trained interviewers who determined eligibility using a short questionnaire. Each interviewer had to include both cases and controls. Blood samples from individuals fulfilling these criteria were collected in ethylene diamine tetra-acetic acid (EDTA) tubes and stored at -20°C. The study population consisted of 150 patients with lung cancer (98 squamous cell carcinomas and 52 small cell carcinomas), 129 patients with cancers of the larynx (55 supraglottic, 47 glottic/subglottic, and 27 unspecified or unclassifiable laryngeal cancers), 121 patients with cancers of the oral cavity/pharynx (67 oral cancers, 50 oro- or hypopharyngeal cancers, and four unspecified or unclassifiable cancers of the oral cavity or pharynx), and 172 control individuals. The main medical diagnoses in the control population were rheumatological (33%) (of which 71% were lumbago and sciatica), infectious and parasitic (10%), respiratory (9%), cardiovascular (8%), digestive (6%) and traumatological (6%) diseases. For the other categories, the main admission reasons were related to general symptoms (7%).

Detailed information on demographic factors, medical history, lifetime tobacco and alcohol use, and occupational exposures was recorded during a personal standardized interview. The daily consumption of each type of tobacco was expressed in g/day (1 g for a cigarette, 2 g for a cigar, and 3 g for a pipe). The average daily consumption of tobacco was calculated by dividing the cumulative lifetime tobacco consumption by the overall duration of smoking. The consumption of alcoholic beverages was expressed in grams of pure ethanol (4.0, 9.4, 14.5 and 31.7g for 0.11 of beer, wine, cider, aperitif and hard liquor, respectively). For the drinkers, the average daily consumption of alcohol was calculated

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Table 1. Main characteristics of the study population.

	Controls (n=172)	Lung cancer (n=150)	Laryngeal cancer (n = 129)	Oral/pharyngeal cancer (n = 121)
Number of males (%) Age (years)	163 (95%)	104 (93%)	127 (98%)	113 (93%)
$Mean \pm SD$	54.9 ± 11.1	58.4 ± 9.9^{a}	55.0 ± 9.4	54.4 ± 10.2
Range	25-88	36-81	22-85	25-89
Mean $(\pm SD)$ tobacco consumption $(g day^{-1})$	25.1 ± 12.5	26.3 ± 13.4	30.4 ± 15.8^{b}	28.2 ± 13.6^a
Mean (±SD) duration of smoking (years)	32.2 ± 11.6	$38.0 \pm 9.4^{\rm c}$	34.6 ± 8.8^a	33.9 ± 10.2
Mean (±SD) alcohol consumption (g day ⁻¹)	77.1 ± 64.0	83.0 ± 80.2	98.1 ± 69.9^{b}	$111.8\pm71.5^{\mathrm{d}}$
Mean (±SD) duration of alcohol drinking (years)	29.0 ± 16.5	29.4 ± 16.5	30.1 ± 14.1	30.3 ± 12.6

^a p < 0.01, t-test (comparison with control individuals).

by dividing the cumulative daily consumption of alcohol (the sum of the number of grams of ethanol per day multiplied by the number of years that the quantity was drunk) by the overall duration of drinking (Péquignot et al. 1988). The main characteristics of the study population are presented in table 1. Since only regular smokers were enrolled, the variability in daily tobacco consumption was, as might have been expected, not markedly different between cases and controls. The daily alcohol intake was significantly higher among patients with cancers of the upper aerodigestive tract than among controls.

Genotyping analyses

DNA was isolated using standard methods and stored at -20°C until use. The GSTM1 genotypes had been analyzed earlier (Jourenkova et al. 1997). The NQO1 genotypes were determined as described previously (Traver et al. 1997). Briefly, the sense (5'-TCCTCAGAGTGGCATTCTGC-3') and antisense (5'-TCTCCTCATCCTGTACCTCT-3') primers were used to amplify over the region of the NQO1 gene comprising the polymorphic site. The HinfI restriction site created by the C⁶⁰⁹T change was used to distinguish between the NQO1 Ser and Pro alleles. Analyses were performed by investigators who were blinded to the subjects' case or control status. To ensure laboratory quality control, two independent readers interpreted the results. Any sample with ambiguous results was retested, and a random selection of 20% of all samples were repeated.

Statistical analyses

Odds ratios (ORs) and 95% confidence intervals (CIs) were calculated by unconditional logistic regression (Breslow and Day 1980). All risk estimates were adjusted for sex, age (< 50, 50-54, 55-59, 60-64 and ≥65 years) and smoking-related variables, i.e. smoking status (ex-smoker/current smoker), inhalation (never/ever), duration of smoking in years (≤25, 26-35 and >35) and daily tobacco consumption in g day⁻¹ (<20, 21-30 and > 30). Multivariate analyses included additional terms for occupational exposures to asbestos (never/ever) or arsenic (never/ever) for lung cancer, and daily consumption of alcohol in $g\,day^{-1}$ (<40, 41–80, 81–120 and >120) for cancers of the upper aerodigestive tract. All of the cut-off points were defined according to the distributions in the control population so that sufficient numbers of individuals were included in each subgroup. Associations between each site of cancer and NQO1 genotype were evaluated by homogeneity tests, and the increase of risk with the number of Ser alleles was tested by linear trend tests (Breslow and Day 1980). Interactions between NQO1 genotype and smoking-related variables were studied to test the equality of the effect of genotype across levels of smoking exposure. These interactive effects were assessed by likelihood ratio tests to compare the goodness of fit of the models with and without the interaction term, taking into account the above-mentioned adjusting factors. For that purpose, the average daily tobacco consumption and duration of smoking were expressed as categorical variables dichotomized at the median in the control population. Because of the small number of individuals homozygous for the NQO1 Ser allele, interaction analyses could not be conducted considering the Ser/Ser genotype separately. Therefore, heterozygous and homozygous NQO1 Ser allele-containing genotypes were RIGHTS LINK()

 $^{^{\}rm b}$ p < 0.01, Wilcoxon's rank-sum test (comparison with control individuals).

 $^{^{\}rm c}$ p < 0.0001, t-test (comparison with control individuals).

 $^{^{\}rm d}$ p=0.0001, Wilcoxon's rank-sum test (comparison with control individuals).

combined. Similar analyses were conducted to test interactions between the NOO1 genotype and GSTM1 genotype (homozygous null versus others).

Results and discussion

The NOO1 genotype distribution in the control population was in Hardy-Weinberg equilibrium (p = 0.24) (table 2). The NQO1 Ser allele was present in 43.3% of lung cancer patients (allele frequency = 0.25), 45.0% of laryngeal cancer patients (allele frequency = 0.24), 40.5% of oral/pharyngeal cancer patients (allele frequency = 0.24) and 39.0% of control individuals (allele frequency = 0.21). The frequency of homozygotes for the Ser allele among lung cancer patients (6.7%) and among oral/pharyngeal cancer patients (7.4%) was twice that among control individuals (2.9%). However, the genotype distribution did not differ significantly between cases and controls (p = 0.26 and p = 0.20, respectively). The adjusted ORs of lung cancer were 1.1 (95% CI 0.7-1.8) for the NQO1 Pro/Ser genotype and 2.2 (95% CI 0.7-6.7) for the Ser/Ser genotype compared with the Pro/Pro genotype ($p_{\text{trend}} = 0.29$). The figures for oral/pharyngeal cancer were 0.8 (95%) CI 0.5-1.5) and 2.3 (95% CI 0.6-8.2), respectively ($p_{trend} = 0.60$). The laryngeal cancer risks associated with NQO1 genotypes were close to 1 (table 2).

We did not find any significant interaction between the NQO1 genotype and either the GSTM1 genotype, the duration of smoking (table 3) or the daily consumption of tobacco (data not shown).

NQO1 has been proposed to play a protective role against the toxic effects of BPQs and to prevent the formation of BPQ-DNA adducts generated during metabolic activation. In this study, the Ser/Ser genotype was associated with a tendency towards a two-fold increased risk for lung cancer and for oral/pharyngeal cancer, but not for laryngeal cancer. These results are consistent with the hypothesis that lack of NQO1 activity may be involved in some smoking-related cancers. However, they were based on small numbers of individuals with the putative at-risk genotype and the associations did not reach statistical significance.

A limitation of our study could be the use of hospital controls, especially if there are any associations between NOO1 genotype and their diseases. However, the genotype distribution was not significantly different among the disease groups, although the power to detect such differences was low. Moreover, the frequencies of the NQO1 Ser allele and of the Ser/Ser genotype in our control population were in agreement with those previously reported in Caucasians (Kelsey et al. 1997, Gaedigk et al. 1998).

To our knowledge, only two previous studies have investigated the association of NQO1 polymorphism with lung cancer risk in Caucasian populations. In one US study (Roswold et al. 1995), the Ser allele was twice as frequent in lung cancer patients, recruited in a cancer centre, than in controls recruited independently of cases (0.22 versus 0.13), but this difference disappeared when cancer patients were compared with employees from the same centre as the cases. Consistent with this, Chen et al. (1999) found no difference in the NQO1 Ser allele frequency between lung cancer patients and controls (0.22 and 0.20, respectively). However, these findings contrast with those observed in some other ethnic populations; a protective effect of the NQO1 Ser allele was found among Mexican-Americans and African-Americans (Wiencke et al. 1997), among Japanese (Chen et al. 1999) and among Taiwanese (Lin et al. 1999). Further studies are therefore clearly needed RIGHTS LINKA)

Table 2. Distribution of individuals by NQOI genotype and ORs (95% CI) of cancer.

	Controls $(n = 172)$	Lur (n	Lung cancer $(n=150)$	Laryn, (n)	Laryngeal cancer $(n=129)$	Oral/phar (n	Oral/pharyngeal cancers $(n = 121)$
$N ilde{Q} O I$ genotype	No. (%)	No. (%)	No. (%) OR^a (95% CI)	No. (%)	No. (%) OR ^b (95%CI)	No. (%)	No. (%) OR ^b (95%CI)
Pro/Pro Pro/Ser Ser/Ser Pro/Ser or Ser/Ser	105 (61.1) 62 (36.1) 5 (2.9) 67 (39.0)	85 (56.7) 55 (36.7) 10 (6.7) 65 (43.3)	35 (36.7) 1 (Reference) 55 (36.7) 1.1 (0.7–1.8) 10 (6.7) 2.2 (0.7–6.7) 55 (43.3) 1.2 (0.7–2.0)	71 (55.0) 54 (41.9) 4 (3.1) 58 (45.0)	1 (Reference) 1.2 (0.7–2.1) 1.1 (0.2–5.9) 1.2 (0.7–2.1)	72 (59.5) 40 (33.1) 9 (7.4) 49 (40.5)	72 (59.5) 1 (Reference) 40 (33.1) 0.8 (0.5-1.5) 9 (7.4) 2.3 (0.6-8.2) 49 (40.5) 0.9 (0.6-1.6)

^aAdjusted for sex, age, smoking and occupational exposures. Data on smoking are missing for four lung cancer cases and three

^bAdjusted for sex, age, smoking exposure and daily consumption of alcohol. Data on smoking and/or alcohol drinking were missing for six laryngeal cancer cases, six oral/pharyngeal cancer cases and eight controls.



Number of cases/controls and ORs (95% CI) of cancer in relation to NQOI genotype by duration of smoking and GSTMI genotype. Table 3.

Duration of smoking \le 30 years No. of cases/controls			•	Tan Jugan amagu	· J have	magnifundhara
Duration of smoking \leq 30 years No. of cases/controls	Pro/Pro	Pro/Pro Pro/Ser or Ser/Ser	Pro/Pro	Pro/Pro Pro/Ser or Ser/Ser	Pro/Pro	Pro/Pro Pro/Ser or Ser/Ser
No. of cases/controls						
	22/51	12/29	23/49	16/28	24/49	17/28
	(Reference)	1.0 (0.4-2.3)	1 (Reference)	1.5 (0.6–3.6)	1 (Reference)	1.3 (0.6–3.0)
Duration of smoking > 30 years						
No. of cases/controls	62/54	52/38	46/50	38/37	46/50	38/37
OR (95% CI) 3.	3.1 (1.4-6.8)	3.7 (1.7–8.2)	2.5 (1.1–5.7)	2.7 (1.2–6.2)	2.0 (0.9-4.3)	1.8 (0.8-4.0)
ntrols	41/47	28/35	29/43	20/33	34/43	25/33
1	(Reference)	1.2 (0.6–2.5)	1 (Reference)	1.2 (0.5–2.7)	1 (Reference)	1.1 (0.5–2.2)
No. of cases/controls	44/58	37/32	40/56	34/32	36/56	20/32
	1.1 (0.6–2.2)	1.4 (0.7–2.8)	1.5 (0.7–3.1)	1.8 (0.9–3.9)	0.9 (0.5–1.8)	0.8 (0.4–1.8)

Interaction test between NQOI genotype and duration of smoking (likelihood ratio test, 1d.f.), p=0.75; interaction test between NQOIgenotype and age, smoking and occupational exposures. Data on smoking are missing for four lung cancer cases and three controls. GSTMI genotype (likelihood ratio test, 1d.f.), p = 0.95. ^aAdjusted for sex,

^bAdjusted for sex, age, smoking exposure and daily consumption of alcohol. Data on smoking and/or alcohol drinking were missing for six aryngeal cancer cases and eight controls. Interaction test between NQOIgenotype and duration of smoking (likelihood ratio test, 1d.f.), p=0.54; interaction test between NQOI genotype and GSTMI genotype (likelihood ratio test, 1d.f.), p = 0.95

^cAdjusted for sex, age, smoking exposure and daily consumption of alcohol. Data on smoking and/or alcohol drinking were missing for six oral/ pharyngeal cancer cases and eight controls. Interaction test between NQOI genotype and duration of smoking (likelihood ratio test, 1d.f.), $\rho = 0.51$; interaction test between NQOI genotype and GSTMI genotype (likelihood ratio test, 1d.f.), p=0.82.



for a better understanding of the potential role of NQO1 activity in tobacco-related cancers.

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References

- BOUCHARDY, C., WIKMAN, H., BENHAMOU, S., HIRVONEN, A., DAYER, P. and HUSGAFVEL-PURSIAINEN, K. 1997, CYP1A1 genetic polymorphisms, tobacco smoking and lung cancer risk in a French Caucasian population. Biomarkers, 2, 131-134.
- Breslow, N. E. and Day, N. E. 1980, The Analysis of Case-Control Studies. Statistical Methods in Cancer Research, Vol. 1, Scientific Publications No 32 (Lyon: International Agency for Research on Cancer).
- Chen, H., Lum, A., Seifried, A., Wilkens, L. R. and Le Marchand, L. 1999, Association of the NAD(P)H:quinone oxidoreductase ⁶⁰⁹C→T polymorphism with decreased lung cancer risk. Cancer Research, 59, 3045-3048.
- Gaedigk, A., Tyndale, R. F., Jurima-Romet, M., Sellers, E. M., Grant, D. M. and Leeder, J. S. 1998, NAD(P)H:quinone oxidoreductase polymorphisms and allele frequencies in Caucasian, Chinese and Canadian native Indian and Inuit populations. Pharmacogenetics, 8, 305-313.
- JOSEPH, P. and JAISWAL, A. K. 1994, NAD(P)H:quinone oxidoreductase 1 (DT-diaphorase) specifically prevents the formation of benzo[a]pyrene quinone-DNA adducts generated by cytochrome P4501A1 and P450 reductase. Proceedings of the National Academy of Sciences of the USA, 91, 8413-8417.
- Jourenkova, N., Reinikanen, M., Bouchardy, C., Husgafvel-Pursiainen, K., Dayer, P., Benhamou, S. and Hirvonen, A. 1997, Effects of glutathione S-transferase GSTM1 and GSTT1 genotypes on lung cancer risk among smokers. Pharmacogenetics, 7, 515-518.
- Jourenkova-Mironova, N., Voho, A., Bouchardy, C., Wikman, H., Dayer, P., Benhamou, S. and HIRVONEN, A. 1999, Glutathione S-transferase GSTM1, GSTM3, GSTP1 and GSTT1 genotypes and the risk of smoking-related oral and pharyngeal cancers. International Journal of Cancer, 81, 44-48.
- Kelsey, K. T., Ross, D., Traver, R. D., Zuo, Z.-F., Spitz, M. R., Wang, M., Xu, X., Lee, B.-K., Schwartz, B. S. and Wiencke, J. K. 1997, Ethnic variation in the prevalence of a common NAD(P)H:quinone oxidoreductase polymorphism and its implications for anti-cancer chemotherapy. British Journal of Cancer, 76, 852-854.
- LIN, P., WANG, H. J., LEE, H., LEE, H. S., WANG, S. L., HSUEH, Y. M., TSAI, K. J. and CHE, C.Y. 1999, NAD(P)H:quinone oxidoreductase polymorphism and lung cancer in Taiwan. Journal of Toxicology and Environmental Health, 58, 187-197. RIGHTSLINK

- Péquignot, G., Crosignani, P., Terracini, B., Ascunce, N., Zubiri, A., Raymond, L., Estéve, J. and Tuyns, A. J. 1988, A comparative study of smoking, drinking, and dietary habits in population samples in France, Italy, Spain, and Switzerland. III. Consumption of alcohol. Revue d'Epidemiologie et de Sante Publique, 36, 177–185.
- Roswold, E. A., McGlynn, K. A., Lustbader, E. D. and Buetow, K. H. 1995, Identification of an NAD(P)H:quinone oxidoreductase polymorphism and its association with lung cancer and smoking. *Pharmacogenetics*, 5, 199–206.
- Siegel, D., McGuinness, S. M., Winski, S. L. and Ross, D. 1999, Genotype-phenotype relationships in studies of a polymorphism in NAD(P)H:quinone oxidoreductase 1. *Pharmacogenetics*, **9**, 113–121.
- TALALAY, P., FAHEY, J. W., HOLTZCLAW, W. D., PRESTERA, T. and ZHANG, Y. 1995, Chemoprotection against cancer by phase 2 enzyme induction. *Toxicology Letters*, 82–83, 173–179.
- Traver, R. D., Horikoshi, T., Danenberg, K. D., Stadlbauer, T. H. W., Danenberg, P. V., Ross, D. and Gibson, N. W. 1992, NAD(P)H:quinone oxidoreductase gene expression in human colon carcinoma cells: characterization of a mutation which modulates DT-diaphorase activity and mitomycin sensitivity. *Cancer Research*, 52, 797–802.
- Traver, R. D., Siegel, D., Beall, H. D., Gibson, N. W., Franklin, W. A. and Ross, D. 1997, Characterization of a polymorphism in NAD(P)H:quinone oxidoreductase (DT-diaphorase). British Journal of Cancer, 75, 69–75.
- VINEIS, P., D'ERRICO, A., MALATS, N. and BOETTA, P. 1999, Overall evaluation and research perspectives. In *Metabolic Polymorphisms and Susceptibility to Cancer*, edited by P. Vineis, N. Malats, M. Lang, A. d'Errico, N. Caporaso, J. Cuzick and P. Boffetta (Lyon: International Agency for Research on Cancer), pp. 403–408.
- WIENCKE, J. K., SPITZ, M. R., McMillan, A. and Kelsey, K. T. 1997, Lung cancer in Mexican-Americans and African-Americans is associated with the wild-type genotype of the NAD(P)H:quinone oxidoreductase polymorphism. *Cancer Epidemiology, Biomarkers and Prevention*, 6, 87–92.
- WORKMAN, P. 1994, Enzyme-directed bioreductive drug development revisited: a commentary on recent progress and future prospects with emphasis on quinone anticancer agents and quinone metabolizing enzymes, particularly DT-diaphorase. *Oncology Research*, **6**, 461–475.

